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# Muscle shear wave elastography, conventional B mode and power doppler ultrasonography in healthy adults and patients with autoimmune inflammatory myopathies: a pilot cross-sectional study



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#### **Abstract**

**Background:** Before the role of shear wave elastography (SWE) and B mode ultrasound (US) in the diagnosis of different forms of idiopathic inflammatory myopathies (IIM) can be investigated, normative data is required. This study aimed to describe and then compare normative SWE and B mode ultrasound metrics of muscles in healthy controls and patients with IIM.

**Methods:** Twenty nine healthy adult controls and 10 IIM patients (5 with inclusion body myositis and 5 with necrotising autoimmune myopathy) underwent a full clinical examination, laboratory investigations, SWE and US measurements of selected proximal and distal limb muscles. Shear wave speed (SWS) and multiple US domains [echogenicity, fascial thickness, muscle bulk and power Doppler (PD)] were measured in both groups.

**Results:** In healthy controls (n = 29; mean age  $46.60 \pm 16.10$ ; 44.8 % female), age was inversely correlated with SWS at the deltoid (stretch) (Rs. -0.40, p = 0.030) and PD score at the deltoid (rest) (Rs. -0.40, P = 0.032). Those ≥ 50 years old had a lower SWS at the deltoid (stretch) compared to the < 50 year group (2.92 m/s vs. 2.40 m/s, P = 0.032). Age correlated with increased echogenicity in the flexor digitorum profundus (Rs. 0.38, P = 0.045). Females had a smaller muscle bulk in the deltoid (P = 0.022). Body mass index (BMI) was inversely associated with SWS in the deltoid (stretch) (Rs − 0.45, P = 0.026), and positively correlated with echogenicity in the deltoid (Rs. 0.69, P = 0.026). In patients ≥50 years of age, patients with IIM (mean age  $61.00 \pm 8.18$ ; females 20.0 %) had a higher proportion of abnormal echogenicity scores at the flexor digitorum profundus (FDP) (40.00 % vs. 14.30 %, P = 0.022) and tibialis anterior (TA) (80.00 % vs. 28.60 %, P = 0.004). Fascial thickness was lower in the FDP (0.63mm vs. 0.50mm, p = 0.012) and TA (0.58mm vs. 0.45mm, P = 0.001).

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**Conclusions:** Our findings suggest there is scope for US techniques to be useful for diagnostic screening of affected muscles in patients with IIM, especially in those with suspected inclusion body myositis or necrotising autoimmune myopathy. We provide normative data for future studies into SWE and US techniques in skeletal muscle. The differences between IIM patients and controls warrant further study in a broader IIM patient cohort.

**Keywords:** Shear wave elastography, ultrasound, idiopathic inflammatory myopathy, myositis, imaging

## **Background**

Idiopathic inflammatory myopathies (IIM) are a rare group of skeletal muscle diseases, unified by immune-mediated muscle damage, that result in muscle fibre loss and weakness. Subtypes of IIM include dermatomyositis (DM), polymyositis (PM), necrotising autoimmune myopathy (NAM), overlap myositis (OM) and inclusion body myositis (IBM) [1]. The subtypes of IIM are heterogeneous with clinically distinct presentations and patterns of muscle involvement, serum autoantibody profiles, and response to therapies. Despite being associated with high morbidity, and increased mortality [2, 3], most subtypes of IIM are treatable; with earlier diagnosis and management usually translating to better health-related outcomes [4].

Currently, muscle biopsy remains the gold standard for confirming the diagnosis of IIM and to exclude other muscle disorders. However, delayed or misdirected biopsies contribute to the 10–45 % false-negative biopsy rate [5]. Increasingly, magnetic resonance imaging (MRI) is used as a screening procedure for the presence and extent of muscle abnormalities, such as inflammation, oedema, fatty infiltration, and muscle atrophy [6]; and may help in determining the optimal site to biopsy [5]. However, the feasibility of MRI is limited by expense, accessibility and patient tolerability.

Point-of-care imaging tools such as shear wave elastography (SWE) and ultrasound (B mode and power Doppler) are being studied to assess their utility in the timelier diagnosis and monitoring of patients with IIM [7–11]. Although these modalities have been evaluated in neuromuscular disease and muscle injury [12], their use in IIM remains understudied, particularly in inclusion body myositis (IBM) and necrotising autoimmune myopathy (NAM) that pose diagnostic and management difficulties for clinicians [13, 14]. Moreover, there are a limited number of studies with normative ultrasound data for muscle groups that are typically most affected by these forms of myopathy[15, 16].

Shear wave elastography (SWE) is a novel ultrasound technique which uses a modified low frequency/high-intensity B mode US push-pulse called, acoustic radiation force impulse (ARFI) to displace tissue and determine muscle stiffness [17, 18]. Muscle stiffness is recorded as Shear Wave Speed (SWS) in m/s, or as Muscle Shear Modulus (MSM), in kilopascals (kPa). Lower SWS is

associated with lower muscle stiffness (10), and SWE has shown age-related differences in muscle stiffness which correlate with muscle weakness [19]. The capability of US to measure tissue perfusion and echogenicity makes it a promising tool to study muscle inflammation [20]. In addition, other US domains findings such as increased fascial thickness and reduced muscle bulk have been described in IIM but are still contentious and have not been validated, largely due to lack of normal values in healthy controls [11] Similarly, published evidence based on the use of SWE suggests that muscle stiffness is reduced with increasing age and in active IIMs, but this needs replication to ensure validity [19].

This study aimed to determine if SWE and US measurements have a role in measuring inflammation or changes in diseased muscle tissue in a group of IIM patients compared to normative data for muscle groups typically affected by the disease. Firstly, this study produces normative metrics with SWE and US of selected muscles in a group of healthy adults, including the influence of age, sex, and BMI. Secondly, this study, evaluated the ability of SWE/US to detect pathology in affected muscles of patients with an established diagnosis of inclusion body myositis (IBM) or necrotising autoimmune myopathy (NAM) by comparing the IIM patients to the healthy controls. We focussed on IBM and NAM because these have been less studied by US/ SWE and because they more often pose problems with confirmation of the diagnosis and treatment than other forms of IIM such as dermatomyositis and overlap syndromes.

# Methods

#### **Subjects**

Participants (≥ 18 years of age) who were willing and able to provide written informed consent, be able to attend a clinical study visit at a single tertiary hospital centre, and attend a separate ultrasound visit at a private imaging centre. IIM patients were eligible for the study if they satisfied the 2017 European League Against Rheumatism (EULAR) classification of IIM and/or 188th Neuromuscular Centre (ENMC) diagnostic criteria for IBM and/or ENMC 2004 diagnostic criteria for immune-mediated NAM [1, 21, 22]. Forty participants, including 30 healthy controls and 10 patients with a biopsy-confirmed IIM were recruited for this study from November 2018 to

March 2019. We excluded one healthy control who failed to attend the ultrasound visit. Two healthy controls were pregnant at the time of the study. Further details of the patient and control groups are provided in Supplementary Table 2. Approval for the study was granted by the South Metropolitan Health Service Human Research Ethics Committee (EC00265). All methods were performed in accordance with the relevant guidelines and regulations.

#### Clinical and laboratory assessments

Data was collected on demographic and clinical characteristics, and patient-reported outcomes, including the health assessment questionnaire (HAQ) [23]; patient global assessment of disease activity visual analogue scale (patient VAS, 0-10) [24]; and physician global assessment of disease activity visual analogue scale (physician VAS, 0-10) [24]. In these three patient-reported outcome scales, higher scores indicate more severe functional impairment or disease activity. All participants underwent a full clinical examination, including manual muscle testing in 26 muscle groups (MMT26) [25]. The MMT26 is a partially validated tool to assess muscle strength at 26 sites using a 0-10-point scale, with higher scores indicating greater muscle strength (maximum score = 260). The MMT26 allows identification of musclespecific strength scores for the deltoid, quadriceps, and the tibialis anterior. Participants also underwent routine haematological and biochemical analyses, including creatine kinase (CK) activity, and myositis autoantibody studies.

#### Imaging Method/landmarks

Imaging was conducted by a sonographer who was blinded to the study groups. As previous studies have shown no significant differences in muscle stiffness (SWS) between dominant and non-dominant limbs [26], imaging of muscle landmarks with SWE, B mode US and power Doppler (PD) were performed on the participant's right-hand side.

# Shear wave elastography

To determine muscle stiffness, SWE (recorded as SWS in m/s) was conducted using a Canon Aplio 500 machine with a linear probe set at 4 MHz frequency. Due to clinic time feasibility, SWE was only assessed in the deltoid and vastus lateralis. These two muscles were chosen because they are commonly involved in IIM [27]. SWS was recorded both at rest and with the muscle passively stretched [6]. For the deltoid, rest was defined as the position in which the elbow was flexed and rested on a pillow, and passive stretch was defined as the position in which the arm hung freely with the elbow extended [6]. In the vastus lateralis, rest was defined as the position in which the knee was extended with the patient supine on a bed [6], while passive stretch was

defined as when the knee was flexed to 45 degrees (Supplementary Figs. 1 and 2).

SWS was chosen over MSM as a measure of muscle stiffness as it is devoid of assumptions that tissue density is constant at 1 g/cm<sup>3</sup> and that all tissues are isotropic and homogenous, which is not the case with muscles [26]. SWS measurement was undertaken with minimal skin pressure, and the probe held in the longitudinal plane to the muscle fibres, which is considered to be the most reliable technique [26]. The probe position for imaging the deltoid was 1/3 of the distance from the acromion to the lateral epicondyle (Supplementary Fig. 1). The vastus lateralis probe position was 1/4 of the distance from the anterior superior iliac spine (ASIS) to the superior border of the patella (Supplementary Fig. 2). In order to determine SWS, two 3mm ROI boxes were placed in a region of uniform elastogram vertical lines aiming for a standard deviation (SD) of less than 20 % of the mean speed (Supplementary Fig. 3) [17]. Care was taken not to place the ROI at the epimysium as it could affect the elasticity [26]. The final SWS value was derived from the average of the two ROIs.

#### B mode ultrasound

B mode US was performed using the same Canon Aplio 500 machine with a 14 – 5 linear probe set at 14 MHz frequency. B mode US imaging was undertaken in the deltoid, flexor digitorum profundus, flexor carpi ulnaris, vastus lateralis, and the tibialis anterior. The probe was placed to capture the short axis image of each muscle, and the deltoid and vastus lateralis were landmarked as described for the SWE measurements. The probe position for the flexor digitorum profundus and flexor carpi ulnaris was 5 cm from the olecranon process (Supplementary Fig. 4). For the tibialis anterior, the probe position was on the anterior shin just proximal to the bulk of the calf (Supplementary Fig. 5).

A semi-quantitative grading of echogenicity was assigned to each muscle group using the Heckmatt visual grading score [28]. Each muscle was assigned a score of 1-4 as a representation of the quantity of echoes displayed in the greyscale image (Figs. 1 and 2) using the echogenicity of cortical bone as a visual anchor [28]. A grade of 4 indicates increased echogenicity and obliteration of cortical bone echo. Fascial thickness (aggregate of dissectible connective tissue sheaths that envelopes muscle) of each muscle was measured in millimetres using the calliper function in a homogenous area of the fascia [29]. Five samples were obtained, each at least 0.25 cm apart, and averaged [30]. Muscle bulk was measured in 3 muscles (deltoid, vastus lateralis and flexor digitorum profundus) by measuring the thickest portion of the muscle belly from the superficial fascia to the deep fascia in cross-section under the predetermined probe positions described above [30].

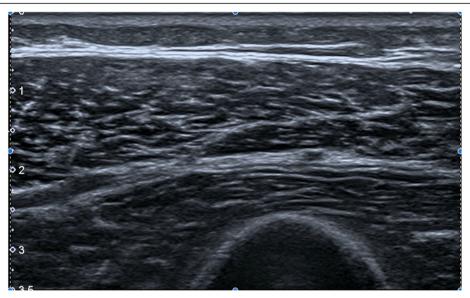


Fig. 1 B mode Ultrasound showing a transverse view of the Vastus Lateralis in a healthy individual

### Power Doppler (PD)

The PD was assessed with a low wall filter and pulse repetition frequency (PRF) at 750 Hz. Colour gain (CG) was increased until the ROI was filled and reduced again until background noise was eliminated. PD images were obtained at rest after B mode images were taken, both in transverse and longitudinal planes. A modified semi-quantitative PD grading scale of 0–4 [31] was used, and the highest PD score was taken as the overall score for each muscle. Grade 0 was defined as no vessels seen; Grade 1: at least one intramuscular vessel seen; Grade 2

was defined as  $\geq 5$  vessels in a two-dimensional frame or a single large intramuscular vessel seen in a cross-section of > 5mm, or a segment length of > 10mm; Grade 3 = vascularity grade 2 with small clusters ( $\geq 3$ ) of vessels; Grade 4 = appearance of frank blush, or vessel boundaries not distinguishable. Higher PD scores indicate greater degrees of vascularity.

The normal ranges for each US domain for each muscle were analysed. As age-related changes in muscle structure and function are known to commence after  $\geq$  50 years of age [32], the healthy control group was



Fig. 2 B mode Ultrasound showing a transverse view of the Vastus Lateralis in a patient with Inclusion Body Myopathy. Muscle echogenicity is increased (appears brighter) and there is loss of muscle bulk

divided into those  $\leq 50$  years of age and those  $\geq 50$  years of age. When comparing with findings in IIM, the healthy control group was restricted to a subgroup comprising of those  $\geq 50$  years of age.

To assess the intra-observer reliability, three months after the initial assessment, a re-reading session was performed with restored images in 21 % of the cases. The analysis of the recorded images was carried out by the first author (SP) who had undergone a previous 6-month period of intensive training under the supervision of an experienced sonographer.

#### Statistical methods

Continuous data are described as either median with interquartile range (IQR) or mean with confidence intervals. Continuous data were compared across IIM patients and healthy controls using the Mann-Whitney U test or One-way ANOVA. Categorical data are described as a frequency with proportion and compared across groups with the Fisher's Exact test. Correlations were done using Spearman's rank correlation coefficient. For statistical comparisons p values < 0.05 were considered significant. For intra-observer comparisons, the ICC was categorised as: slight agreement (0.00-0.20), fair agreement (0.21–0.40), moderate agreement (0.41–0.60), substantial agreement (0.61–0.80), and almost perfect agreement (0.81-1.00) [33]. Statistical analysis was carried out using SPSS version 24.0 (IBM Corp. in Armonk, NY).

#### **Results**

#### Normative muscle metrics in controls

Complete data for shear wave speed (muscle stiffness), echogenicity, fascial thickness, muscle bulk, and power Doppler scores (vascularity) was available for each muscle examined in 29 healthy controls (mean age 46.60 ± 16.1; female 44.80 %) (Table 1). There were no statistically significant differences between the values across the two age groups (< 50 years and ≥50 years), except that SWS in the deltoid at passive stretch was significantly lower in the ≥50 year group compared to the < 50 year group (2.80 vs. 2.47, p = 0.033). Comparison of the metrics for the individual US domains showed variation between different muscle groups. For example, SWS measure of muscle stiffness was greater in the deltoid than in the vastus lateralis and was higher in the stretched than in the rest state. Echogenicity and power Doppler values varied over a broad range (1-3 and 0-4 respectively) in the muscles examined. In this study, echogenicity of > 1 is likely outside of the normal range in all muscles except the vastus lateralis where the threshold was > 2. Fascial thickness was greater in the proximal limb muscles (deltoid, vastus lateralis) than in the distal muscles (flexor digitorum profundus, flexor carpi ulnaris, tibialis anterior).

## Influence of age, sex and BMI on normative muscle metrics

The influence of age, sex and BMI on healthy muscle is presented in Supplementary Table 1. Age was found to have significant correlations with a number of US domains. SWS in the deltoid at passive stretch correlated inversely with age (Rs: -0.40, P = 0.030). Echogenicity in the flexor digitorum profundus was positively correlated with age (Rs. 0.38, P = 0.045). Similarly, PD was positively correlated with age in the deltoid (P = 0.032). Females had lower muscle bulk in the deltoid (P = 0.022). No gender influences were noted for SWS, echogenicity or PD. Higher BMI was associated with lower SWS (Rs. -0.45, P = 0.018) and higher echogenicity (Rs. 0.69, P < 0.001) in the deltoid, but was not associated with PD score in any muscle group (P > 0.10).

# Comparison of muscle metrics in IBM/NAM cases and healthy controls $\geq$ 50 years

Muscle imaging findings were compared in 13 healthy controls  $\geq 50$  years (mean age  $61.00 \pm 7.47$ ) and 10 participants with IIM (mean age 67.00+/-8.18 years); 5 with IBM and 5 with NAM). There were more males in the IIM group (n = 8/10, 80.0 %) and more females in the healthy control  $\geq 50$  years group (n = 8/14, 57.1%), which was not statistically significant (P = 0.069). Participants with IIM recorded a lower MMT26 (i.e., weaker muscles), a higher participant and physician global activity VAS (i.e., more disease activity), HAQ scores (i.e., more disability), and a higher average serum CK level, these differences all being statistically significant (p < 0.002). IBM patients had a lower BMI and CK levels compared to NAM, and higher participant and physician VAS and higher HAQ scores, which were all statistically significant (p < 0.049) (Supplementary Table 2).

In-group comparisons for the deltoid muscle between the five NAM cases and healthy controls  $\geq$  50 years, no significant differences were apparent in SWS (P = 0.099), fascial thickness (P = 0.220), muscle bulk (P = 0.727), echogenicity (P = 0.199) or PD score (P = 0.086) (Supplementary Table 3). However, fascial thickness was significantly reduced in the tibialis anterior (P = 0.002) and flexor digitorum profundus (P = 0.004). Analysis of SWS values on a case-to-case basis showed that values in all five cases were lower than the mean for the and healthy controls  $\geq$  50 years group, particularly in the stretched muscle condition (Table 2), and SWS values were also lower in the vastus lateralis in a number of cases (Table 2).

In IBM, the findings in the flexor digitorum profundus, tibialis anterior and vastus lateralis muscles were compared with controls, based on the known predilection of IBM for these muscles. In the flexor digitorum profundus, echogenicity was increased (P = 0.022) and fascial thickness was reduced (P = 0.04), while PD scores (P = 0.881) did not differ significantly between the two

**Table 1** Descriptive data in the healthy control group showing normal ranges for each ultrasound domain in each muscle examined, presented as median, interguartile range, minimum and maximum values

	Median (IQR), Min-Max	p- value <sup>a</sup> (comparing medians)		
	Age < 50	Age ≥50		
Fascial Thickness (mm)				
Deltoid	0.76(0.66-0.84)0.50-0.92	0.72(0.64-0.76)0.48-0.90	0.123	
Vastus Lateralis	0.80(0.68-0.92)0.64 -1.00	0.86(0.70-0.93)0.50-1.00	0.627	
Tibialis Anterior	0.59(0.50-0.70)0.48-0.72	0.61(0.56-0.71)0.52-0.72	0.477	
Flexor Digitorum Profundus	0.54(0.48-0.63)0.42-0.70	0.59(0.52-0.64)0.46-0.68	0.329	
Flexor Carpi Ulnaris	0.60(0.56-0.64)0.36-0.74		0.451	
Muscle Bulk (mm)				
Deltoid	19.80(14.40-22.20)9.40-27.3	17.30(15.88–19.35)11.4–27.1	0.354	
Vastus Lateralis	20.15(17.20-2150)13.2-25.3	17.60(14.28-22.65)13.1-24.4	0.377	
Flexor Digitorum Profundus	10.55(9.20-11.30)8.30-15.2	12.60(9.28-14.90)8.00-17.1	0.252	
Shear Wave Speed (m/s)				
Deltoid (rest)	2.31(1.94–2.55)1.58–2.83	2.14(1.79-2.47)1.36-2.85	0.652	
Deltoid (Stretch)	2.80(2.37-3.24)1.65-4.68	2.47(2.18-2.63)1.72-2.97	0.333	
Vastus Lateralis (Rest)	1.70(1.63-1.75)1.59-3.34	1.74(1.57-1.97)1.34-2.08	0.747	
Vastus Lateralis (Stretch)	1.79(1.70-1.97)1.47-3.08	1.90(1.81-2.04)1.62-2.43	0.270	
Echogenicity (1-4)			P value <sup>b</sup> (comparing medians)	
Deltoid	1(1,1)1–3	1 (1,1)1–3	0.553	
Vastus Lateralis	1(1, 1)1–2	1(1-2)1-3	0.451	
Flexor digitorum profundus	1(1,1) 1–1	1 (1,1) 1–2	0.533	
Flexor carpi ulnaris	1(1-1)1-1	1 (1,1) 1-1	0.747	
Tibialis anterior	1(1-1)1-2	1 (1,2)1–2	0.331	
Power Doppler (0-4)			P value <sup>b</sup> (comparing medians	
Deltoid	0 (0-1) 0-2	0 (0,0) 0–2	0.134	
Vastus Lateralis	1 (0,1) 0-1	1(0–1) 0–2 0.949		
Flexor Digitorum Profundus	0 (0,1) 0–2	1(0-1) 0-2	0.477	
Flexor Carpi Ulnaris	1 (0,1) 0-3	1(0-1) 0-4	0.533	
Tibialis Anterior	1 (0,1) 0-1	1(0-1) 0-2	0.252	

IQR interquartile range, min-max: minimum-maximum, P value<sup>a</sup>: Mann-Whitney test, P value<sup>b</sup>: Fisher Exact Test

groups. In the tibialis anterior, echogenicity was increased (P = 0.004), fascial thickness was significantly reduced (P = 0.002), while PD scores did not differ between groups (P = 0.518). In the vastus lateralis muscle bulk was significantly reduced (P = 0.033). Mean SWS values in the vastus lateralis and deltoid did not differ significantly from the healthy controls  $\geq$  50 years group (supplementary Table 3), but case-to-case analysis showed that in a number of cases, SWS values were lower than the mean for the healthy controls  $\geq$  50 years group in the resting state, and higher in the stretched state (Table 2).

#### Intra-observer reliability

The images of seven healthy controls and one patient with IIM were reviewed for reliability. Intra-observer variability

for scoring echogenicity on B mode ultrasound demonstrated perfect intraclass correlation coefficient agreement (kappa 1.00) [33] in all of the deltoid, vastus lateralis, flexor digitorum profundus, flexor carpi ulnaris and tibialis anterior muscles. PD scoring showed substantial agreement in vastus lateralis (kappa 0.85), flexor digitorum profundus (kappa 0.87), deltoid (kappa 0.67) and moderate agreement in the flexor carpi ulnaris (kappa 0.43).

#### **Discussion**

#### Normative data

Although SWE, B mode US and PD have been studied previously in patients with musculoskeletal disorders (6, 26, 29, 30), to our knowledge, this is the first study in which they have been used in combination in a group of healthy controls to establish normal metrics and US

Table 2 Case-by-case analysis of shear wave speed (SWS) in the IBM and NAM groups with corresponding age and muscle weakness

Study Subtype	Age	SWSD Rest-m/s	SWSD Stretch-m/s	SWSVL Rest-m/s	SWSVL Stretch-m/s	MMTD /10	MMT quads/10
NAM	69	1.75	2.04	1.95	2.04	9	10
NAM	71	1.75	1.79	1.89	2.87	9	8
NAM	52	1.77	2.03	1.68	1.70	6	7
NAM	56	2.67	2.29	1.67	1.81	10	8
NAM	70	2.17	2.12	1.55	2.28	7	8
IBM	76	1.78	4.05	1.83	4.19	7	5
IBM	77	2.56	2.10	1.50	1.77	10	9
IBM	68	1.61	2.51	1.41	3.43	10	7
IBM	61	1.81	2.25	1.71	2.30	5	9
IBM	70	1.66	2.59	2.16	1.30	10	7
HC Mean+/-SD	61.00, 7.47	2.14+/-0.43	2.40+/-0.36	1.74+/-0.24	1.93+/-0.20	10	10

D deltoid, VL vastus lateralis, quads quadriceps femoris, SWS shear wave speed, m/s metres per second, MMT manual muscle testing (maximum score 10), NAM necrotising autoimmune myopathy, IBM inclusion body myositis, HC healthy controls, SD standard deviation

profiles for selected upper and lower limb muscles which are typically affected in IIM. Previous studies of SWE and ultrasound in immune-mediated IIMs have focussed on dermatomyositis, polymyositis and overlap syndromes, and there is a paucity of studies of IBM and the more recently characterised subtype of NAM. The primary objective was to evaluate the properties of healthy muscle with SWE, B mode US and PD, and to undertake an exploratory assessment of how they are altered in a small group of patients with IBM and NAM which have not been studied previously. Normative data in the adult population are lacking and most previous studies have focussed on the paediatric population which may have different muscle metrics. In our study, healthy controls generally showed low grades of echogenicity and PD in all muscles examined with an age influence of echogenicity in the flexor digitorum profundus and power Doppler at the deltoid.). We found that echogenicity > 1 is likely to be of significance in all muscles studied, except the vastus lateralis which had a higher threshold of > 2. The increased threshold in the quadriceps may be due to the accelerated sarcopenic effects in the quadriceps muscles and increasing myosteatosis compared to its counterparts [19, 34]. A PD score > 1 was also likely significant in all muscles, but it is unclear whether this is necessarily pathological as there may be other influences on muscle vascularity such as exercise, temperature, medications and hyperdynamic states that need further study.

#### Influences of age, sex and BMI on healthy muscle

Previous published data have indicated that age may be an influence on muscle ultrasound properties [35, 36], especially in men [34]. Therefore, we examined the impact of age on healthy controls in this study. Our SWE data in healthy controls indicated an age-associated decline in

deltoid SWS at passive stretch but not at rest, indicating a reduction in muscle stiffness. As muscles at rest and stretch may have different physical properties [37], SWS should therefore be considered contextually with regards to whether the muscle is stretched or not [37] [15]. The decline in SWS in the deltoid with age in healthy controls has been reported by Alfuraih et al. but in contrast to our findings, the age-associated decline was seen at rest and not in the passively stretched state [19]. We found no age-related changes in SWE stiffness in the vastus lateralis, in contrast to the study by Alfuraih et al. which showed a decline in stiffness in the vastus lateralis at rest with age. This difference may be related to differences in sample size, age distributions, and different limb positions during the procedure and definitions of stretch and rest.

In our study, BMI was found to influence muscle stiffness and echogenicity in the deltoid. Higher echogenicity and lower muscle stiffness were associated with a higher BMI in the deltoid. The association of higher BMI and higher echogenicity is expected [38] but the predilection to the deltoid is interesting and warrants further study. Apart from the expected greater muscle bulk in the deltoid in males, there were no gender-related differences in muscle stiffness or other US measures in other muscles.

# Comparison between healthy controls $\geq$ 50 years and IIM muscles

In this preliminary work considering healthy controls  $\geq 50$  years, compared to IIM participants, differential changes in SWE muscle stiffness were found in different muscles in patients with NAM and IBM. In NAM, SWS was reduced relative to controls in both the stretched deltoid and vastus lateralis, which are both proximal limb muscles affected by the disease, but the group differences were not statistically significant, most likely because of the small patient

numbers. In IBM, the group changes in SWS in the vastus lateralis, which is one of the muscles most severely affected by the disease [39] were not significantly different from healthy controls  $\geq 50$  years. However, there was a greater variability in SWS than in controls in both the resting and stretched muscle conditions. Our preliminary findings suggest that there are changes in the physical properties of affected muscles in these forms of IIM, but these require further investigation in larger patient cohorts in which correlations with parameters of disease severity and duration can be investigated. SWE findings of reduced muscle stiffness in IIM compared to healthy controls has been demonstrated in a previous study, by Alfuraih et al.[6, 19], but the study did not include patients with NAM or IBM.

Other changes in the muscle properties in the IIM group included increased echogenicity, which was present in almost all muscle groups examined compared to healthy controls ≥ 50 years. In the acute setting, increased echogenicity may be the result of inflammation or fatty replacement (myosteatosis) [11, 40]. However, the people with IIM in this study were all prevalent cases, without clinical evidence of inflammation, and therefore the increased echogenicity in these chronically affected muscles is likely to be a result of myosteatosis in which there is progressive loss of muscle fibres, as is known to occur in IBM [11, 41]. It is thought that fatty replacement increases echogenicity as opposed to intramuscular fibrosis [42]. However, the US correlation with interstitial muscle fibrosis has not been clearly defined and further studies that involve comparison with pathology would be needed to evaluate this.

Another finding of note was a reduction in fascial thickness in the flexor digitorum profundus and tibialis anterior muscles in IIM compared to healthy controls. To date, published data regarding fascial thickness has focused on the IIM subsets of DM and PM, and has described an increase in fascial thickness [43], which is in contrast to our findings in NAM and IBM. We are unaware of any previous published data on fascial thickness in NAM and IBM and the factors responsible for a reduction in fascial thickness are unclear. The increased fascial thickness in DM is thought to occur as a result of vascular endothelial growth factor (VEGF) expressing cells and angiogenesis in the fascia (37). However, given VEGF is not a prominent cytokine in IBM and NAM, it is intuitive that fascial thickness would not be increased in IBM and NAM [44]. The reason for the facial thinning is not clear; the role of prednisolone and the underlying disease pathology on fascia in these subtypes is unknown and may have an influence on the fascia.

No differences in PD signal were seen between healthy controls and people with IIM. A single previous published study reported increased PD signal in acute IIM [45]. A few possibilities could explain this discordance. The healthy control group in the present study reported higher exercise intensity on history (compared to IIM),

which could influence muscle vascularity. Exercise has been reported to increase intramuscular vascularity as perfusion increases [31]. Additionally, there were two patients in our study who were pregnant. We are unable to find published data to demonstrate that pregnancy, a hyperdynamic state, definitively increases vascularity in muscle, but it remains a possibility. The chosen PD grading score, which includes the presence of large intramuscular vessels, (as opposed to just fine tissue vascularity) may also have been a factor. Finally, in contrast to the published study (37) which examined acute, incident IIM (although exclusive of NAM), our cohort was a prevalent group so that muscle inflammation may have been reduced as a result of treatment.

#### Limitations

This study has a number of limitations. Firstly, as SWE and US are operator-dependent, they can be affected by subjective changes in the degree of pressure applied to the muscle being examined. However, care was taken to minimise this by using a standardised predefined and previously validated protocol for each muscle examined. Secondly, the study included only patients with NAM and IBM and did not include other forms of IIM such as dermatomyositis, polymyositis or overlap syndromes. Thirdly, as an exploratory pilot study, the number of subjects included in the IIM group was small (n = 10), and the study was not sufficiently powered for subgroup analysis or correlations with effects of treatment or with parameters of disease activity or clinical outcome measures. Overall, while a high degree of inter-observer reliability and high ICCs was demonstrated in the analysis of ultrasound images, the possibility that the retrospective analysis of stored images may have affected the ICCs for the power doppler analysis cannot be excluded.

#### **Conclusions**

This study provides normative SWE and US data for healthy controls at sites clinically relevant to IBM and NAM. We have shown influence of age and BMI on SWE and US properties in the deltoid which is an important consideration when choosing a muscle to study. Future work investigating whether ultrasound has the potential to be a clinically useful tool should examine its performance when compared to current diagnostic and clinical outcome measures with particular attention to construct validity, longitudinal discrimination and feasibility.

#### **Abbreviations**

IIM: Aidiopathic inflammatory myopathies; SWE: Shear wave elastography; MSM: Muscle shear modulus; SWS: Shear wave speed; PD: Power Doppler; DM: Dermatomyositis; PM: Polymyositis; IBM: Inclusion body myositis; NAM: Necrotising autoimmune myositis; D: Deltoid; FDP: Flexor digitorum profundus; FCU: Flexor carpi ulnaris; VL: Vastus lateralis; TA: Tibialis anterior;

BMI: Body mass index; MMT26: Manual muscle testing for 26 muscle; VAS: Visual analogue scale; HAQ: Health assessment questionnaire

#### **Supplementary Information**

The online version contains supplementary material available at https://doi.org/10.1186/s12891-021-04424-0.

**Additional file 1: Supplementary Table 1**: Age, gender and BMI influences in healthy controls.

**Additional file 2: Supplementary Table 2:** Baseline characteristics of IIM patients and subgroup of healthy controls over 50 years of age.

**Additional file 3: Supplementary Table 3:** Discrimination of ultrasound domains between IIM and healthy controls.

**Additional file 4: Supplementary Figure 1**: The Deltoid probe position was 1/3 of the distance from the acromion to the lateral epicondyle.

**Additional file 5: Supplementary Figure 2**: The Vastus Lateralis probe position was ¼ of the distance from the anterior superior iliac spine (ASIS) to the superior border of the patella.

**Additional file 6: Supplementary Figure 3**: Shear wave speed in the deltoid of a 28-year-old healthy female at rest showing the colour blue, which indicates lower muscle stiffness.

**Additional file 7: Supplementary Figure 4:** The Flexor Digitorium Profundus and Flexor Carpi Ulnaris probe position was 5 cm from olecranon process.

**Additional file 8: Supplementary Figure 5:** The Tibialis Anterior muscle probe position is anterior shin just proximal to the calf bulk.

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#### Authors' contributions

The corresponding author (SP), the senior author (HK) and second author (MN) were responsible for obtaining the grant, protocol development, ethics approval, recruitment, data collection. The corresponding author (SP) and co-author (WR) are responsible for the statistics. Co-author (DL) is a research assistant who helped with recruitment and data collection. Co-author (NM) is a sonographer who helped with the technical aspects of the US, and SWE data analysis and the co-author (PC) helped with the protocol development. The co-author (FM) is a neurologist who has helped with the presentation of the data. All co-authors were involved in the manuscript creation and revisions. The authors read and approved the final manuscript.

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#### Availability of data and materials

All data generated or analysed during this study are included in this published article [and its supplementary information files.

#### **Declarations**

#### Ethics approval and consent to participants

Approval for the study was granted by the South Metropolitan Health Service Human Research Ethics Committee (EC00265). Authors SP (first author), MN (second author) and HIK (senior author) secondary affiliation is with Fiona Stanley Hospital which comes under South Metropolitan Health Service which is the primary ethics committee and the primary site for recruitment. We have also received reciprocal ethics approval from Murdoch University (Permit number: 2018/237), grounds on which the neuromuscular research clinic was in.All participants have given a written informed consent

#### Consent for publications

N/A

#### Competing interests

All patients had no competing interest.

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#### References

- Bottai M, Tjarnlund A, Santoni G, Werth VP, Pilkington C, de Visser M, Alfredsson L, Amato AA, Barohn RJ, Liang MH, et al. EULAR/ACR classification criteria for adult and juvenile idiopathic inflammatory myopathies and their major subgroups: a methodology report. RMD Open. 2017;3(2):e000507.
- Limaye VS, Lester S, Blumbergs P, Roberts-Thomson PJ. Idiopathic inflammatory myositis is associated with a high incidence of hypertension and diabetes mellitus. Int J Rheum Dis. 2010;13(2):132–7.
- Furst DE, Amato AA, lorga SR, Bancroft T, Fernandes AW. Medical costs and health-care resource use in patients with inflammatory myopathies in an insured population. Muscle Nerve. 2012;46(4):496–505.
- Malik A, Hayat G, Kalia JS, Guzman MA. Idiopathic Inflammatory Myopathies: Clinical Approach and Management. Front Neurol. 2016;7:64.
- Day J, Patel S, Limaye V. The role of magnetic resonance imaging techniques in evaluation and management of the idiopathic inflammatory myopathies. Semin Arthritis Rheum. 2017;46(5):642–9.
- Alfuraih AM, O'Connor P, Tan AL, Hensor EMA, Ladas A, Emery P, Wakefield RJ. Muscle shear wave elastography in idiopathic inflammatory myopathies: a case-control study with MRI correlation. Skeletal Radiol. 2019. https://doi. org/10.1007/s00256-019-03175-3.
- Alfuraih AM, O'Connor P, Tan AL, Hensor E, Emery P, Wakefield RJ. An investigation into the variability between different shear wave elastography systems in muscle. Med Ultrason. 2017;19(4):392–400.
- Youk JH, Son EJ, Gweon HM, Kim H, Park YJ, Kim JA. Comparison of strain and shear wave elastography for the differentiation of benign from malignant breast lesions, combined with B-mode ultrasonography: qualitative and quantitative assessments. Ultrasound Med Biol. 2014;40(10): 2336–44.
- Kim HJ, Lee HK, Cho JH, Yang HJ. Quantitative comparison of transient elastography (TE), shear wave elastography (SWE) and liver biopsy results of patients with chronic liver disease. J Phys Ther Sci. 2015;27(8):2465–8.
- Tavare AN, Alfuraih AM, Hensor EMA, Astrinakis E, Gupta H, Robinson P. Shear-Wave Elastography of Benign versus Malignant Musculoskeletal Soft-Tissue Masses: Comparison with Conventional US and MRI. Radiology. 2019; 290(2):410–7.
- Paramalingam S, Morgan K, Becce F, Diederichsen LP, Ikeda K, Mandl P, Ohrndorf S, Sedie AD, Sharp V, Tan AL, et al. Conventional ultrasound and elastography as imaging outcome tools in autoimmune myositis: A systematic review by the OMERACT ultrasound group. Semin Arthritis Rheum. 2020. https://doi.org/10.1016/j.semarthrit.2020.11.001.
- Pillen S, Arts IM, Zwarts MJ. Muscle ultrasound in neuromuscular disorders. Muscle Nerve. 2008;37(6):679–93.
- 3. Liang C, Needham M. Necrotizing autoimmune myopathy. Curr Opin Rheumatol. 2011;23(6):612–9.
- Needham M, Mastaglia FL. Sporadic inclusion body myositis: A review of recent clinical advances and current approaches to diagnosis and treatment. Clin Neurophysiol. 2016;127(3):1764–73.
- Botanlioglu H, Kantarci F, Kaynak G, Unal Y, Ertan S, Aydingoz O, Erginer R, Unlu MC, Mihmanli I, Babacan M. Shear wave elastography properties of

- vastus lateralis and vastus medialis obliquus muscles in normal subjects and female patients with patellofemoral pain syndrome. Skeletal Radiol. 2013; 42(5):659–66.
- Wang L, Xiang X, Zhu BH, Qiu L. Determination of reference ranges for normal upper trapezius elasticity during different shoulder abduction using shear wave elastography: a preliminary study. Sci Rep. 2020;10(1):17104.
- O'Hara S, Zelesco M, Rocke K, Stevenson G, Sun Z. Reliability Indicators for 2-Dimensional Shear Wave Elastography. J Ultrasound Med. 2019;38(11): 3065–71.
- Taljanovic MS, Gimber LH, Becker GW, Latt LD, Klauser AS, Melville DM, Gao L, Witte RS. Shear-Wave Elastography: Basic Physics and Musculoskeletal Applications. Radiographics. 2017;37(3):855–70.
- Alfuraih AM, Tan AL, O'Connor P, Emery P, Wakefield RJ. The effect of ageing on shear wave elastography muscle stiffness in adults. Aging Clin Exp Res. 2019. https://doi.org/10.1007/s40520-019-01139-0.
- Terslev L, Naredo E, Keen HI, Bruyn GAW, Iagnocco A, Wakefield RJ, Conaghan PG, Maxwell LJ, Beaton DE, Boers M et al: The OMERACT Stepwise Approach to Select and Develop Imaging Outcome Measurement Instruments: The Musculoskeletal Ultrasound Example. J Rheumatol. 2019. https://doi.org/10.3899/jrheum.181158
- Rose MR, Group ElW: 188th ENMC International Workshop: Inclusion Body Myositis, 2–4 December 2011, Naarden, The Netherlands. Neuromuscul Disord. 2013, 23(12):1044–1055.
- Hoogendijk JE, Amato AA, Lecky BR, Choy EH, Lundberg IE, Rose MR, Vencovsky J, de Visser M, Hughes RA: 119th ENMC international workshop: trial design in adult idiopathic inflammatory myopathies, with the exception of inclusion body myositis, 10–12 October 2003, Naarden, The Netherlands. Neuromuscul Disord. 2004, 14(5):337–345.
- Bruce B, Fries JF. The Health Assessment Questionnaire (HAQ). Clin Exp Rheumatol. 2005;23(5 Suppl 39):S14-18.
- 24. Ruperto N, Ravelli A, Pistorio A, Ferriani V, Calvo I, Ganser G, Brunner J, Dannecker G, Silva CA, Stanevicha V, et al. The provisional Paediatric Rheumatology International Trials Organisation/American College of Rheumatology/European League Against Rheumatism Disease activity core set for the evaluation of response to therapy in juvenile dermatomyositis: a prospective validation study. Arthritis Rheum. 2008;59(1):4–13.
- Rider LG, Giannini EH, Harris-Love M, Joe G, Isenberg D, Pilkington C, Lachenbruch PA, Miller FW. International Myositis A, Clinical Studies G: Defining Clinical Improvement in Adult and Juvenile Myositis. J Rheumatol. 2003;30(3):603–17.
- Alfuraih AM, O'Connor P, Hensor E, Tan AL, Emery P, Wakefield RJ. The
  effect of unit, depth, and probe load on the reliability of muscle shear wave
  elastography: Variables affecting reliability of SWE. J Clin Ultrasound. 2018;
  46(2):108–15.
- Joyce NC, Oskarsson B, Jin LW. Muscle biopsy evaluation in neuromuscular disorders. Phys Med Rehabil Clin N Am. 2012;23(3):609–31.
- 28. Heckmatt JZ, Leeman S, Dubowitz V. Ultrasound imaging in the diagnosis of muscle disease. J Pediatr. 1982;101(5):656–60.
- Stecco C, Tiengo C, Stecco A, Porzionato A, Macchi V, Stern R, De Caro R. Fascia redefined: anatomical features and technical relevance in fascial flap surgery. Surg Radiol Anat. 2013;35(5):369–76.
- Bhansing KJ, van Rosmalen MH, van Engelen BG, van Riel PL, Pillen S, Vonk MC. Muscle ultrasonography is a potential tool for detecting fasciitis in dermatomyositis and polymyositis: comment on the article by Yoshida. Arthritis Rheumatol. 2017;69(11):2248–9.
- Meng C, Adler R, Peterson M, Kagen L. Combined use of power Doppler and gray-scale sonography: a new technique for the assessment of inflammatory myopathy. J Rheumatol. 2001;28(6):1271–82.
- Faulkner JA, Larkin LM, Claflin DR, Brooks SV. Age-related changes in the structure and function of skeletal muscles. Clin Exp Pharmacol Physiol. 2007; 34(11):1091–6.
- Landis JR, Koch GG. An application of hierarchical kappa-type statistics in the assessment of majority agreement among multiple observers. Biometrics. 1977;33(2):363–74.
- Akima H, Yoshiko A, Tomita A, Ando R, Saito A, Ogawa M, Kondo S, Tanaka NI. Relationship between quadriceps echo intensity and functional and morphological characteristics in older men and women. Arch Gerontol Geriatr. 2017;70:105–11.
- Arts IM, Pillen S, Schelhaas HJ, Overeem S, Zwarts MJ. Normal values for quantitative muscle ultrasonography in adults. Muscle Nerve. 2010;41(1):32–41.

- Maurits NM, Bollen AE, Windhausen A, De Jager AE, Van Der Hoeven JH. Muscle ultrasound analysis: normal values and differentiation between myopathies and neuropathies. Ultrasound Med Biol. 2003;29(2):215–25.
- Dubois G, Kheireddine W, Vergari C, Bonneau D, Thoreux P, Rouch P, Tanter M, Gennisson JL, Skalli W. Reliable protocol for shear wave elastography of lower limb muscles at rest and during passive stretching. Ultrasound Med Biol. 2015;41(9):2284–91.
- Pereira AZ, Uezima CB, Zanella MT, Prado RRD, Gonzalez MC, Zheng J, Heymsfield SB. Muscle Echogenicity and Changes Related, to Age and Body Mass Index. JPEN J Parenter Enteral Nutr. 2020. https://doi.org/10.1002/ ioen.2030.
- Needham M, James I, Corbett A, Day T, Christiansen F, Phillips B, Mastaglia FL. Sporadic inclusion body myositis: phenotypic variability and influence of HLA-DR3 in a cohort of 57 Australian cases. J Neurol Neurosurg Psychiatry. 2008;79(9):1056–60.
- 40. Young HJ, Jenkins NT, Zhao Q, McCully KK. Measurement of intramuscular fat by muscle echo intensity. Muscle Nerve. 2015;52(6):963–71.
- 41. Zaidman CM, van Alfen N. Ultrasound in the Assessment of Myopathic Disorders. J Clin Neurophysiol. 2016;33(2):103–11.
- Reimers CD, Fleckenstein JL, Witt TN, Muller-Felber W, Pongratz DE. Muscular ultrasound in idiopathic inflammatory myopathies of adults. J Neurol Sci. 1993;116(1):82–92.
- 43. Yoshida K, Ito H, Furuya K, Ukichi T, Noda K, Kurosaka D. Angiogenesis and VEGF-expressing cells are identified predominantly in the fascia rather than in the muscle during the early phase of dermatomyositis. Arthritis Res Ther. 2017;19(1):272.
- 44. Dalakas MC. Pathogenesis of immune-mediated neuropathies. Biochim Biophys Acta. 2015;1852(4):658–66.
- Sousa Neves J, Santos Faria D, Cerqueira M, Afonso MC, Teixeira F. Relevance of ultrasonography in assessing disease activity in patients with idiopathic inflammatory myopathies. Int J Rheum Dis. 2018;21(1):233–9.

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