

RESEARCH ARTICLE

Open Access

The development of whole blood titanium levels after instrumented spinal fusion – Is there a correlation between the number of fused segments and titanium levels?

Ingmar Ipach^{1*}, Ralf Schäfer¹, Falk Mittag¹, Carmen Leichtle¹, Petra Wolf² and Torsten Kluba¹

Abstract

Background: Most modern spinal implants contain titanium and remain in the patient's body permanently. Local and systemic effects such as tissue necrosis, osteolysis and malignant cell transformation caused by implants have been described. Increasing tissue concentration and whole blood levels of ions are necessary before a disease caused by a contaminant develops. The aim of the present study was the measurement of whole blood titanium levels and the evaluation of a possible correlation between these changes and the number of fused segments.

Methods: A prospective study was designed to determine changes in whole blood titanium levels after spinal fusion and to analyze the correlation to the number of pedicle screws, cross connectors and interbody devices implanted.

Blood samples were taken preoperatively in group I (n = 15), on the first, second and 10^{th} day postoperatively, as well as 3 and 12 months after surgery.

Group II (n = 16) served as a control group of volunteers who did not have any metal implants in the body. Blood samples were taken once in this group.

The number of screw-rod-connections and the length of the spinal fusion were determined using radiographic pictures. This study was checked and approved by the ethical committee of the University of Tuebingen.

Results: The mean age in group I was 47 ± 22 years (range 16 - 85 years). There were three male (20%) and twelve female (80%) patients. The median number of fused segments was 5 (range 1 to 11 segments).

No statistically significant increase in the titanium level was seen 12 months after surgery (mean difference: $-7.2 \mu g/l$, 95% CI: $-26.9 \text{ to } 12.5 \mu g/l$, p = 0.446). By observing the individual titanium levels, 4 out of 15 patients demonstrated an increase in titanium levels 12 months after surgery.

No statistically significant correlation between fused segments (r = -0.188, p = 0.503) length of instrumentation (r = -0.329, p = 0.231), number of interbody devices (r = -0.202, p = 0.291) and increase of titanium levels over the observation period was seen.

Conclusions: Instrumented spinal fusion does not lead to a statistically significant increase in whole blood titanium levels. There seems to be no correlation between the number of pedicle screws, cross connectors and interbody devices implanted and the increase of serum titanium levels.

Keywords: Spinal fusion, Titanium serum levels, Interbody devices, Pedicle screws

Tuebingen, Germany

Full list of author information is available at the end of the article



^{*} Correspondence: Ingmar.lpach@med.uni-tuebingen.de

¹Department of Orthopaedic Surgery, University Hospital Tuebingen,

Background

Spinal surgery -especially instrumented spinal fusion-has increasing importance in treatment of patients with chronic low back pain caused by osteoligamentary instability, degenerative disc diseases or spondylolisthesis. Most of the modern spinal implants contain titanium and remain in the patient's body permanently. Little is known about the influence of free metal ions on a patient's health. Negative effects such as tissue necrosis, osteolysis and malignant cell transformation have been described [1].

Rising tissue concentration and serum levels of ions are necessary before a disease caused by a contaminant develops [2,3]. It has been shown that metal ions such as vanadium and cobalt chromium decrease monocyte and macrophage survival in a dose-dependent manner [4].

It has also been shown that titanium-alloy particles stimulate cytokine release. Additionally opsonization of machined particles with human serum increased the release of macrophages and cytokines [5].

Activation of the monocyte-macrophage-system leads to a local inflammatory reaction and is a relevant promoter for prosthetic loosening [6].

Although titanium is characterized by excellent qualities, there are many indications that it is not inert. Nephrotoxicity, cardiac and hepatic injury have been described after oral administration to rats [7]. Patients undergoing revision for non-fusion after posterior lumbar spondylodesis, were observed to have a high

paraspinal tissue concentration of titanium [8], and rising serum titanium levels after spinal fusion with a correlation to the amount of fused segments has been demonstrated [9]. Free titanium particles lead to an increase in prostaglandin and interleukin synthesis as an indicator of an inflammatory process with a negative influence on bone formation [10,11].

Corrosion of titanium ions in the surrounding tissue and the movement of these ions into the distant organs may cause systemic problems. By using rat peritoneal macrophages and by determining metal concentration in sheep after spinal fusion, an accumulation in the different organs especially in the brain, lungs, liver, spleen, kidneys and lymph nodes was detected. Inflammation and osteolysis was seen in the macrophages [12-16]. On the other hand, titanium is present as a micronutrient in the body even without any metal implants. A normal value for titanium in blood seems to be between 50-150 ug/l (Synlab/Leinfelden, Germany).

The aim of the present study was to detect a possible increase in titanium levels after spinal fusion. We also wanted to investigate a possible correlation between the number of implanted pedicle screws, cross connectors and interbody devices on titanium levels.

Methods

A prospective study was performed to detect the effects of spinal fusion on titanium levels. For instrumentation, the Muenster-Anterior/Posterior-Doublerod-

Table 1 Demographic data group I

Patient	sex	age	Diagnosis	Fused levels	length of intstrumantation	Number of interbody devices	Type of Implant	Titanium levels (ug/l)				
						,	•	Präoperativ	2. p.o.	10. p.o.	3 Mon p.o.	12 Mon p.o.
1	f	73	DLS	10	1626	3	MPDS	56	34	26	29	37
2	f	55	LS	1	381	1	MPDS	33	44	Ø	63	36
3	f	16	Scoliosis	6	864	0	MADS	34	Ø	48	Ø	31
4	f	16	Scoliosis	6	869	0	MADS	39	43	39	Ø	96
5	f	22	Scoliosis	5	811	0	MADS	48	Ø	42	54	94
6	m	45	Scoliosis	5	1076	2	MADS	54	42	44	55	31
7	f	68	LS	1	370	1	Expedium	53	Ø	39	54	33
8	f	85	LSCS	4	824	1	Expedium	61	49	49	63	96
9	f	61	LS	1	410	1	Expedium	53	Ø	49	40	47
10	f	61	LSCS	2	550	1	Expedium	49	Ø	31	45	35
11	m	30	LS	1	405	1	Expedium	47	52	Ø	40	142
12	f	42	Scoliosis	6	965	0	MADS	38	Ø	Ø	37	42
13	f	20	Scoliosis	11	1520	0	Expedium	51	42	3	38	37
14	f	55	DLS	6	1415	0	Expedium	69	33	37	32	36
15	m	58	DLS	4	540	0	Expedium	34	34	45	82	34

DLS: Degenerativ lumbar scoliosis, LS: lumbar spondylolisthesis, LSCS: lumbar spinal canal stenosis, MAPS: Muenster-Anterior-doublerod-system, MPDS: Muenster-posterior-doublerod-system.

System[®] (Schäfer. Micomed, Schorndorf, Germany) and the Expedium[®] (Depuy-spine Raynham, Massachusetts, USA) systems were used.

Group I patients (n = 15) underwent instrumented spinal fusion in a period between January 2006 and March 2009. Blood samples were taken preoperatively, on the first, second and $10^{\rm th}$ day postoperatively, as well as 3 and 12 months after surgery (Table 1).

To evaluate a possible influence of lifetime exposure on titanium levels, a control group (group II) was created. Group II (n=16) consisted of volunteers who did not have any metal implants in the body and were statistically significantly younger than group I. Blood samples were taken once in this group (Table 2).

Only patients with a normal liver and kidney function and no previous history of metal implants were included in the present study.

The number of screw-rod-connections and the length of the spinal fusion were determined using radiographic pictures (Figure 1, 2).

Study registration and informed consent

This study was checked and approved by the local ethical committee of the University of Tuebingen. Written informed consent was obtained from all patients to take part in this study.

Blood samples and titanium analysis

Blood samples were taken by using titanium-free canulas and EDTA-tubes.

Table 2 Demographic data group II

Patient	sex	age	Titanium levels (ug/
1	f	26	59
2	m	55	47
3	m	33	30
4	f	24	37
5	f	55	33
6	m	24	35
7	m	31	57
8	f	50	59
9	f	28	58
10	f	29	55
11	f	50	56
12	m	29	61
13	f	24	33
14	f	24	53
15	f	20	58
16	m	30	59

Collected blood was stored at 5°C until measurement. Titanium was measured by ICP-MS (inductively coupled plasma mass spectrometry) (Synlab/Leinfelden, Germany). Therefore titanium was ionized through plasma at a temperature of 5000°C. Depending on their mass, ions were separated and counted using a spectrometer. Detection limit of titanium ions was >0.02 parts per trillion.

Normal values for whole blood titanium levels were in a range between 50-150 ug/l in blood-samples.

Statistical analysis

Categorical variables are presented as frequencies and percentages, and continuous variables as means and standard deviations, or medians and ranges. A paired *t*-test was used to evaluate the difference in titanium levels preoperatively and 12 months after surgery. To compare the two groups, independent sample t tests or exact Mann Whitney *U* test were used. Differences are presented with 95% confidence intervals (CI). A relationship of titanium levels with fused segments, length of instrumentation and number of interbody devices was evaluated using the Spearman correlation coefficient.

All reported P values are two-sided, with a significance level of 0.05 and have not been adjusted for multiple testing. All analyses were performed with the use of R 2.13.2 (R Foundation for Statistical Computing, Vienna, Austria) and IBM SPSS 20.

Results

The mean age in group I was 47 ± 22 years (range 16 - 85 years). There were three male (20%) and twelve female (80%) patients. The median number of fused segments was 5 (range 1 to 11 segments). Demographic data of this group are shown in Table 1.

The whole blood titanium levels in group I before and in the postoperative period are shown in Table 3.

No statistically significant increase in titanium levels was seen 12 months after surgery (mean difference: -7.2 µg/l, 95% CI: -26.9 to 12.5 µg/l, p = 0.446) (Figure 3). 4 out of 15 patients demonstrated an increase in titanium levels 12 months after surgery (ID 4,5,8,11) (Figure 4). 8 out of 15 patients presented with subnormal levels of titanium.

Titanium levels were evaluated in all patients before surgery and 12 months after surgery. Titanium levels were evaluated in 9 patients (out of 15) two days after surgery, 10 days after surgery in 12 patients (out of 15) and 3 months after surgery in 13 patients (out of 15). 11 patients presented sub-normal titanium levels12 months after surgery.

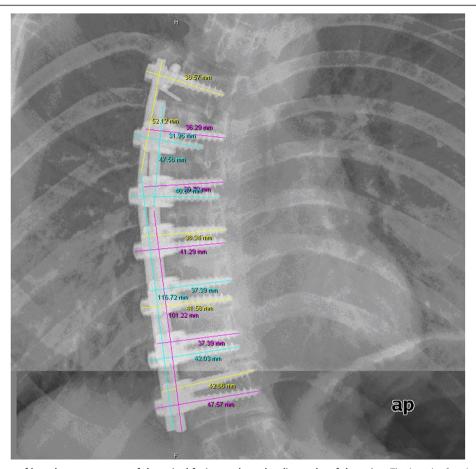


Figure 1 Technique of length-measurement of the spinal fusion on lateral radiographs of the spine. The length of each screw and rod were added to obtain the total length of spinal fusion.

One patient had to be revised due to a loss of correction 6 months after primary surgery, and reinstrumentation was necessary. The development of titanium levels of this patient is also shown in Table 1 (ID 1).

The mean age in group II was 33 ± 12 years (range 20-55 years). There were 6 male and 10 female patients in this group. The mean titanium level in this group was $49.4\pm11.5~\mu g/l$ (range $30-61~\mu g/l$). There was a statistically significant difference regarding age between the two groups (mean difference: 14 years, 95% CI: 0.5 to $27.2~\nu ears$, p = 0.042).

When comparing preoperative titanium levels in group I with group II, there was no statistically significant difference (p = 0.464).

No statistically significant correlation between the number of fused segments (r = -0.188, p = 0.503), length of instrumentation (r = -0.329, p = 0.231), number of interbody devices (r = -0.202, p = 0.291) and development of titanium levels over the observation period was seen.

Discussion

Titanium as a light and strong material is often used as an alloy in orthopedic implants, and is generally regarded as safe for the organism.

Titanium is present as a micronutrient in the body even without metal implants, and most is stored intracellularly. We therefore analyzed not only serum titanium levels as done in previous studies [9,17], but the total amount of titanium in the blood.

Titanium alloy used in pedicle screws and interbody devices contained approximately 90% titanium, 6% aluminum and 4% vanadium [18-20]. Therefore an evaluation of titanium levels in patients after instrumented spinal fusion might be interesting with regard to possible effects caused by metal particles and for early detection of none-fusions.

In the present study, no statistically significant raise in titanium levels during the observation period after spinal fusion was seen. In contrast to our findings, previous studies demonstrated an increase in titanium levels [9,17]. One possible explanation for the different

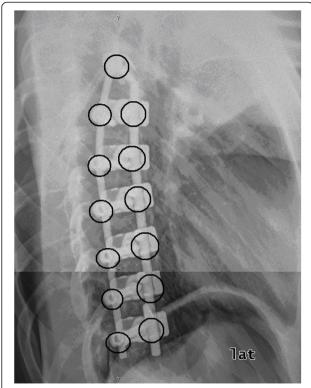


Figure 2 Technique for counting the different connections on a.p. radiographs of the spine.

results might be the use of different samples for the evaluation of titanium levels. As mentioned above, we used whole blood samples in the present study as most titanium is present in macrophages. Travis et al. [9] and Kasai et al. [17] analyzed free titanium ions in serum.

Interestingly, when observing the individual development of whole blood titanium levels, 4 out of 15 patients demonstrated an increase in titanium levels 12 months after surgery. It is unclear why there was such a spread in titanium levels 12 months after surgery. The distribution of metal ions after spinal fusion seems to be a multifactorial process which has not been evaluated. As we analyzed whole blood samples, the total amount of titanium was evaluated

(free and intracellular titanium ions). These findings support previous findings by Richardson et at [9] who also stated that the individual's biological response and elimination of titanium ions might be extremely variable. Another possible explanation might be that these four individuals have undergone unknown environmental exposure. Another possible reason for the increase in titanium levels in these individuals might be a reduced renal clearance, but all patients demonstrated normal renal function during the observation period. Individual titanium elimination might also explain why 8 out of 15 patients presented with sub-normal levels of titanium before surgery, and 10 out of 15 patients 12 months after surgery.

Initial titanium levels seem to have no effect on postoperative titanium levels. The fact that the probands in group II were younger than the patients in group I does not support the theory of continuously rising of levels during a lifetime caused by a longer environmental exposure time.

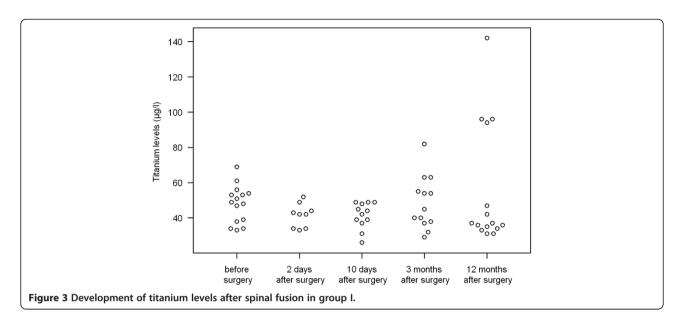
There was no correlation between the number of pedicle screws, cross connectors and interbody devices which support the findings by Travis et al. [9] who also found no correlation. One possible explanation for this finding might be that there is a balance between absorption from the alloy and renal elimination.

One of the main strengths of the present study is the use of whole blood samples. To our knowledge, this is the first study evaluating titanium levels after spinal fusion in whole blood. As demonstrated by Mohammed Bakir [16], there is strong contact between monocytes and the titanium surface with a possible absorption of titanium particles. This might explain the different standard values for titanium in serum (<7 ug/l) and whole blood 50-150 ug/l). By using whole blood samples, total amount of titanium and not only free ions could be detected.

We also note several limitations of the present study. First, the whole titanium levels at the second and 10th postoperative day are incomplete, caused by a sample error. The results for this point of time might therefore not be meaningful.

Table 3 Whole blood titanium levels in group I before and in the postoperative period

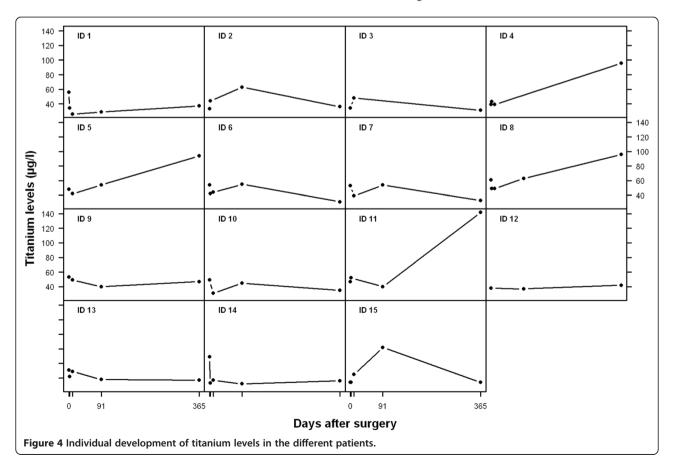
time	mean titanium level (µg/l)	n	standard deviation	minimum titanium level (μg/l)	maximum titanium level (μg/l)	
before surgery	47.9	15	10.6	33.0	69.0	
2 th day after surgery	41.4	9	6.7	33.0	52.0	
10 th day after surgery	41.5	12	7.5	26.0	49.0	
3 months after surgery	48.6	13	15.0	29.0	82.0	
12 months after surgery	55.1	15	34.4	31.0	142.0	



We did not differentiate between the two spinal implant systems used when analyzing serum titanium levels. This might also be of interest and should be evaluated in further studies. Because of the small sample numbers, there may be low power with a possible type II error.

Conclusion

Finally, there seems to be no increase in serum titanium levels after spinal fusion. Whether the trend is stable, or if whole titanium levels increase in patients with non-fusion or after a period of 12 months needs to be investigated in further observations.



Competing interests

The authors declare that they have no competing interests.

Authors' contributions

II: participated in the design of the study, drafted the manuscript, RS: collecting data, FM: participated in the of the study, collecting data, CL: performed spinal surgery, PW: performed statistical analysis, TK: conceived of the study, and participated in its design and coordination. All authors read and approved the final manuscript.

Funding

No funding has been received for this work.

Author details

¹Department of Orthopaedic Surgery, University Hospital Tuebingen, Tuebingen, Germany. ²Technical University of Munich, Department of Medical Statistics and Epidemiology, Ismaninger Str. 22, Munich 81675, Germany.

Received: 20 December 2011 Accepted: 20 August 2012 Published: 27 August 2012

References

- Cobb AG, Schmalzreid TP: The clinical significance of metal ion release from cobalt chromium metal-on-metal hip joint arthroplasty. Proc Instit Mech Eng 2006, 220(2):385–398.
- Savarino L, Greco M, Cenni E, et al: Differences in ion release after ceramic-on-ceramic and metal-on-metal total hip replacement. Mediumterm follow-up. J Bone Joint Surg Br 2006, 88(4):472–476.
- Brodner W, Bitzan P, Meisinger V, et al: Serum cobalt levels after metalon-metal total hip arthroplasty. Bone Joint Surg Am 2003, 85-A(11):2168–2173.
- Lacey DC, De Kok B, Clanchy FI, et al: Low dose metal particles can induce monocyte/macrophage survival. J Orthop Res 2009, 27(11):1481–1486.
- Maloney WJ, Sun DH, Nakashima Y, et al: Effects of serum protein opsonization on cytokine release by titanium-alloy particles. J Biomed Mater Res 1998, 41(3):371.
- Flecher X, Rolland C, Rixrath E, et al: Local and Systemic Activation of the Mononuclear Phagocyte System in Aseptic Loosening of Total Hip Arthroplasty. J Clin Immunol 2009, 29(5):681–690.
- Wang J, Zhou G, Chen C, et al: Acute toxicity and biodistribution of different sized titanium dioxide particles in mice after oral administration. Toxicol Lett 2007, 68(2):176–185.
- Wang JC, Yu WD, Sandhu HS, et al: Metal debris from titanium spinal implants. Spine 1999, 24:899–903.
- Richardson TD, Pineda SJ, Strenge KB, Van Fleet TA, MacGregor M, Milbrandt JC, Espinosa JA, Freitag P: Serum titanium levels after instrumented spinal arthrodesis. Spine 2008, 33:792–796.
- 10. Tsutsui T, Kawaguchi H, Fujino A, et al: Exposure of macrophage-like cells to titanium particles does not affect bone resorption, but inhibits bone formation. J Orthop Sci 1999, 4:32–38.
- Granchi D, Verri E, Ciapetti G, et al: Bone resorbing cytokines in serum of patients with aseptic loosening of hip protheses. J Bone Jonit Surg Br 1998, 80:912–917.
- Haynes DR, Rogers SD, Hay S, et al: The differences in toxicity and release of bone resorbing mediators induced by titanium and cobaldchromium-alloy wear particles. J Bone Joint Surg Am 1993, 75:825–833.
- Brayda-Bruno M, Fini M, Pierini G, et al: Evaluation of systemic metaldiffusion after spinal pedicular fixation with titanium alloy and stainless steel system: a 36-months experimental study in sheep. Int Artif Organs 2001, 24:41–49.
- Krupa D, Baszkiewicz J, Kozubowski JA, et al: Effect of calcium ion implantation on the corrosion resistance and biocompatibility of titanium. Biomaterilas 2001, 22:2139–2151.
- Nagagawa M, Matsuya S, Udoh K: Corrosion behaviour of pure titanium and titanium alloys in fluoride-containing solutions. Dent Mater J 2001, 20:305–314.
- Bakir M: Haemocompatibility of titanium and its alloys. J Biomater Appl 2012. 27:3–15.
- Kasai Y, Iida R, Uchida A: Metal concentrations in the serum and hair of patients with titanium alloy spinal implants. Spine 2003, 28:1320–1326.

- Banovetz JM, Sharp R, Probe RA, et al: Titanium plate fixation: a review of implant failures. J Orthop Trauma 1996, 10:389–394.
- Head WC, Bauk DJ, Emerson RH Jr: Titanium as the material of choice for cementless femoral components in total hip arthroplasty. Clin Orthop 1995. 311:85–90.
- Rocca M, Fini M, Giaversi G, et al: Tibial implants: biochemical and histomorphometric studies of hydroxyapatit-coated and uncoated stainless steel and titanium screws in long-term ovariectomized sheep. Int J Artif Organs 2001, 24:649–654.

doi:10.1186/1471-2474-13-159

Cite this article as: lpach et al.: The development of whole blood titanium levels after instrumented spinal fusion – Is there a correlation between the number of fused segments and titanium levels? BMC Musculoskeletal Disorders 2012 13:159.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

